



Case Study

BQUA PLUS Validation for Commissioning Testing of Ballast Water Management Systems

Overview

Ballast tanks within ships are filled with water to provide stability during transportation of cargo. The ballast water is drawn up from the ships departure location (uptake), and later released at its destination port (discharged). The ballast water that is discharged at the ship’s destination may carry invasive species or pathogens that can disrupt the ecology of delicate marine environments and have substantial environmental and economic impacts. For this reason, the International Maritime Organization (IMO) introduced a Ballast Water Management Convention treaty to implement standards and compliance for ballast water management. The IMO standard D-2 outlines the maximum number of viable organisms allowed to be discharged (Table 1) per volume of ballast water in each of the sample fractions based on the size of organisms. This standard will need to be met by all ships by 2024, hence the need for ballast water management systems (BWMS) to decrease biomass to compliant levels.

Table 1: D-2 Standard Specifications for ballast water discharge compliance.

Organism Size Fractions	D-2 Control Limit	Units
≥ 50 µm	< 10	viable organisms/m ³
≥ 10 µm <50 µm	< 10	viable organisms/mL
Toxicogenic <i>Vibrio Cholerae</i>	<1	colony-forming unit (CFU)/100mL
<i>Escherichia coli</i>	<250	CFU/100mL
Intestinal <i>Enterococci</i>	<100	CFU/100mL

Adenosine triphosphate (ATP) is present in all living cells acting as an energy transporter in metabolic processes. Since it is found in all living cells, it can be used as a true indicator of living organisms in a sample. The LuminUltra **BQUA PLUS Ballast Water Monitoring Kit** is specialized for measurement of ATP in ballast water to provide clear indications of discharge compliance in the three organism size fractions in the D-2 Standard. The BQUA PLUS kit is accurate, rapid, and portable, allowing it to be used on board vessels or in a laboratory.

In this study, the BQUA PLUS kit was assessed by Ankron Water Services GmbH, a marine testing and consulting company. It was tested during a commissioning testing study for samples with simulated BWMS discharge water using chlorine and UV treatment. An indicative analysis using a compliance monitoring device (CMD) is required to validate the commissioning of a BWMS as per IMO regulation BWM.2/Circ.70/Rev.1. The commissioning test is successful if the discharge samples are compliant with the D-2 Standard according to the indicative analysis method used, which are listed in BWM.2/Circ.42/Rev.2. If the samples are non-compliant after treatment, it is recommended to investigate through a detailed analysis. The corresponding ATP concentration guidelines for the D-2 control limits are specified in Table 2.

Table 2: LuminUltra compliance guidelines for ballast water based on ATP measurements.

Fraction	ATP Concentration			
	Unit	Most Likely Compliant	Signal Close to Limit	Most Likely Not Compliant
≥50 µm	pg/m ³	< 150,000	150,000 to 750,000	>750,000
≥10<50 µm	pg/mL	<500	500-1,500	>1,500
Bacteria	pg/100 mL	<1,000	1,000-5,000	>5,000

Methodology



I. Samples

The samples used in this study were artificial marine water spiked above the D-2 discharge limits with organisms that correspond to each size fraction so the BWMS efficacy can be assessed. Copepods (zooplankton) were added to the artificial marine water to make up the $\geq 50 \mu\text{m}$ fraction, and Tetraselmis (phytoplankton) were used for the $\geq 10 < 50 \mu\text{m}$ fraction (Figure 1). The phytoplankton was added to the $\geq 10 < 50 \mu\text{m}$ fraction to 10^4 cells/mL. For both BWMSs being tested, an untreated artificial uptake sample was compared to treated artificial discharge samples.

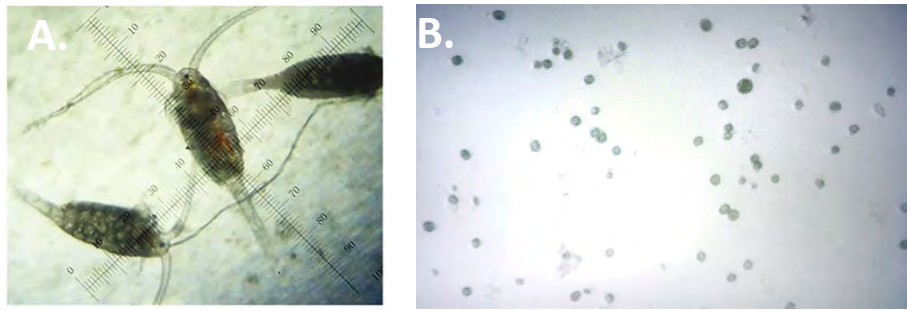


Figure 1: Copepods (A) and Tetraselmis (B) under microscope.

Source: Ankron Water Services GmbH, Dr. Claudia Dreszer, 2021

II. Analysis Methods

The analysis methods used to quantify organisms in the $\geq 10 < 50 \mu\text{m}$ fraction included two detailed (MPN, FDA/CMFDA) and two indicative (PAM, ATP) methods:

- **Most Probable Number (MPN):** MPN is a growth-based method that determines the viable cell count. MPN will not capture cells that are viable but not culturable since these cells will not grow in the nutrient medium.
- **Fluorescein diacetate (FDA)/5-Chloromethylfluorescein diacetate (CMFDA):** The FDA/CMFDA dyes react with enzymes associated with substrate metabolism to emit fluorescence. The sample can then be placed under a fluorescent microscope and the stained cells can be counted. Cells with damaged DNA (non-viable) may be labelled as live cells using this method since their enzymes may still be active.
- **Pulse Amplitude Modulation (PAM):** PAM detects photosynthetic microorganisms by measuring chlorophyll concentration and can give an insight on the biological activity of phytoplankton.
- **ATP measurements:** The BQUA PLUS kit uses the luciferase enzyme isolated from fireflies which emits light upon reacting with ATP. The light is then measured in a luminometer to quantify ATP in the sample.

To quantify the $\geq 50 \mu\text{m}$ fraction, one detailed (microscopy) and one indicative (ATP) method was applied.

- **Microscopy:** direct count of organisms that show any signs of movement or activity.
- **ATP measurements:** Application of the BQUA PLUS kit.

III. Ballast Water Management Systems (BWMS)

Under the chlorine BWMS, the spiked artificial marine water was subject to a chlorine dosage of 8 ppm with a 24-hour holding time. A UV BWMS was simulated using a Collimated Beam Device, where the spiked artificial marine water was subjected to a UV dose of 1.200 J/m^2 before and after a 24-hour holding time for a total UV dose of 2400 J/m^2 . Both the chlorine and the UV dosages used in this study are standard for ballast water management using these systems.



Results and Discussion

The BQUA PLUS ATP data for the chlorine BWMS agreed with the microscopy quantification for the $\geq 50 \mu\text{m}$ fraction, reading well below the D-2 compliance threshold of $150,000 \text{ pg/m}^3$ for this fraction after chlorine treatment (Table 3). Similarly, the ATP data for the $\geq 10 < 50 \mu\text{m}$ fraction agreed with PAM, MPN and FDA/CMFDA methods in measuring a compliant number of organisms after treatment compared to the pre-chlorinated uptake. The commissioning testing of the chlorine BWTS was validated for both the $\geq 50 \mu\text{m}$ and $\geq 10 < 50 \mu\text{m}$ size fractions by the BQUA PLUS ATP test kit and agreed with the other CMD and detailed analyses.

Table 3: ATP measurements of two size fractions of ballast water ($\geq 50 \mu\text{m}$ and $\geq 10 < 50 \mu\text{m}$) compared with detailed analyses and one other CMD before and after chlorine treatment at 8 ppm for a 24-hour holding time. Red shading indicates non-compliance with the D-2 Standard, and green indicates compliance.

	Copepods $\geq 50 \mu\text{m}$		Tetraselmis $\geq 10 < 50 \mu\text{m}$				
	Microscopy	BQUA PLUS ATP Test Kit	PAM		MPN	FDA/CMFDA	BQUA PLUS ATP Test Kit
	org/m ³	cATP ₅₀ pg/m ³	Fv/Fm	biological activity	org/mL	org/mL	cATP ₁₀₋₅₀ pg/mL
Pre-Chlorination	599	1185292	0.6	Healthy	>11000	11427	16313
Post-Chlorination (8 ppm)	0	627	0	Dead	0	0.7	11

The BQUA PLUS ATP data for the $\geq 50 \mu\text{m}$ fraction of the UV treatment agreed with the microscopy results indicating non-compliance (Table 4). In the $\geq 10 < 50 \mu\text{m}$ fraction, the ATP data agreed with the MPN results in showing compliance post-UV treatment compared to the pre-UV treated uptake sample. The PAM and FDA/CMFDA results, however, indicated non-compliance. The PAM and FDA/CMFDA results are likely false positives due to their detection of chlorophyll and metabolic products which can both remain in cells after being rendered non-viable by the UV treatment. The MPN and BQUA PLUS ATP methods were not impacted due to their detection of viable cells only and reported compliance after UV treatment.

Table 4: ATP measurements of two size fractions of ballast water ($\geq 50 \mu\text{m}$ and $\geq 10 < 50 \mu\text{m}$) compared with detailed analyses and one other CMD before and after UV treatment. The total UV dosage for both Low and Medium Pressure UV was 2400 J/m^3 , dosed before and after a 24-hour holding time. Red shading indicates non-compliance with the D-2 Standard, and green indicates compliance.

	Copepods $\geq 50 \mu\text{m}$		Tetraselmis $\geq 10 < 50 \mu\text{m}$				
	Microscopy	BQUA PLUS ATP Test Kit	PAM		MPN	FDA/CMFDA	BQUA PLUS ATP Test Kit
	org/m ³	cATP ₅₀ pg/m ³	Fv/Fm	biological activity	org/mL	org/mL	cATP ₁₀₋₅₀ pg/mL
Pre-UV Treatment	1133	21585540	0.5	Healthy	867	30667	69714
Post-UV Low Pressure (Total dose 2400 J/m^3)	1125	8070900	0.5	Healthy	0.08	288	419
Post-UV Medium Pressure (Total dose 2400 J/m^3)	485	3855610	0.3	Not optimal conditions	0	56	201



Conclusion

Compared with other indicative compliance monitoring tools, the BQUA PLUS test kit that measures ATP concentration of live cells is one of the only tests capable of measuring all three of the D-2 standard size fractions of ballast water: $\geq 50 \mu\text{m}$, $\geq 10 < 50 \mu\text{m}$, and $< 10 \mu\text{m}$ (bacteria). This study demonstrates the suitability of the BQUA PLUS kit as an indicative analysis tool for commissioning testing of ballast water management systems, particularly in its accurate assessment of the $\geq 10 < 50 \mu\text{m}$ fraction after UV treatment. In addition to its ability to accurately detect live cells, the BQUA PLUS ATP assay gives conclusive and rapid results for indicative testing, allowing Ankron Water Services GmbH and other companies performing commissioning testing on BWM systems and avoiding the cost and time needed for excessive detailed analyses.

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